

Original Research

The Effect of the Human Plasma Molecule GHK-Cu on Stem Cell Actions and Expression of Relevant Genes

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Abstract

Background: Stem cell technology is a promising research area with a potential to create effective therapies for many degenerative diseases. However, to apply stem cell technology, we need to be able to identify and understand mechanisms that distinguish healthy regeneration processes from processes, which may result in chronic inflammation, scarring, fibrosis or cancer. GHK-Cu (glycine-L-histidine-lysine) is a small copper-binding peptide, which has a remarkable and well-documented ability to improve wound healing and tissue regeneration, regulate remodeling of connective tissue and synthesis of collagen, elastin and glycosaminoglycans, reduce inflammation and scarring, increase antioxidant-enzymes and protect cells from toxic by-products of lipid peroxidation.

Methods: Authors used a computer-based gene profiling tool, The Connectivity Map, to identify a number of human genes regulated by GHK, relevant to regulation of cell differentiation, apoptosis and stem cell function.

Results: The number of human genes associated with stem cell function was 57 genes in the range of increases of 50% UP and 46 genes in the range of decreases of 50% DOWN.

Conclusion: Based on laboratory data and gene profiling data, GHK-Cu may be used to improve stem cell therapy and to help shift regeneration processes to healthy regeneration.



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Keywords

GHK-Cu; stem cells; matrikines; wound healing; tissue regeneration; gene profiling

1. Introduction

With senior populations growing in many countries, stem cells attract more and more attention as a promising resource in anti-aging therapies. Their ability to repair tissues, which plays a key role in restoring an organ's functionality after an injury, puts them in the forefront of current research in regenerative medicine.

There are three main types of stem cells – embryonic stem cells, adult stem cells and cancer stem cell-like cells. Embryonic stem cells are true omnipotent stem cells, which makes them ideal candidates for regenerative medicine treatments; however, their use is hindered by ethical considerations. Therefore, understanding adult stem cells and harnessing their regenerative potential have become important goals in current regenerative medicine research [1].

A promising source of adult stem cells for stem cell therapy is from epidermal stem cells residing in skin. As recent studies indicate, epidermal stem cells' behavior depends on their niche. When skin is intact, resident epidermal stem cells are committed to their differentiation programs. However, when skin is injured, stem cells are able to migrate from their niche and change their differentiation program, acquiring a high degree of plasticity [2].

Another branch of current research is focused on stem cell-like cells found in certain types of cancer, which are believed to be the cells actively involved in metastasis, tumor invasion and cancer relapse. The mechanisms by which cancer growth can induce stemness in senescent cells are currently investigated [3].

Up until recently, it was believed that cellular senescence, which slows down and stops cell division may be a protective mechanism against cancer. However, a surprising discovery revealed that cellular senescence may elicit stemness, and that this mechanism plays an important role in cancer relapse [4, 5].

It is increasingly apparent that tissue regeneration is a process that shares many mechanisms with cancer growth. In order to restore the tissue's integrity, not only do resident stem cells have to be activated, but also some differentiated cells must revert to a stem cell-like state and acquire mobility. In inflammation and cancer growth, certain inflammatory mediators such as IL 6 and IL 8, can induce stemness in senescent cells [6]. Some pathways that are activated during the wound healing process and inflammation can lead to an increase in cancer stem cell-like population. So, molecules that reduce inflammation and improve regulation of the cytokines and pathways in the course of the wound healing process can help prevent cancer growth [7].

There is increasing understanding of the complexity of how genes regulate functions of stem cells [8]. Regeneration often is accompanied by inflammation and reorganization of the tissue architecture. In order to do this, certain genes that previously were silent, or down-expressed, have to be activated, while others have to be silenced. Today, researchers have an unprecedented opportunity to access gene profiling databases and determine which genes are affected during various physiological and pathological processes. These databases, such as The Broad Institute

Connectivity Map, allow researchers to analyze gene expression changes caused by various biologically active molecules [9].

Also, it is now discovered that inflammation and the accompanying generation of free radicals of oxygen (ROS) can trigger tissue plasticity by reverting cells to pluripotency [10]. Studies show that inflammation in the prostate may expand the pool of progenitor-like stem cells that are prone to malignant transformation [11].

Therefore, one of the primary focuses of regenerative medicine research is identifying and understanding mechanisms that distinguish healthy regeneration processes, which lead to restoration of normal tissue architecture, and pathological processes, which may result in chronic inflammation, scarring, fibrosis or cancer. One of the mechanisms that are currently investigated is the Wnt signalling pathway. A normally functioning Wnt signalling pathway ensures tissue regeneration and maintenance of tissue integrity, while deregulated Wnt plays a role in aging and cancer [12]. Another key player is the p53 gene, which is now recognized as a cancer suppressor and important regulator of microenvironment in the stem cell niche [13].

It is established that stem cells are influenced by their niche and take cues from their microenvironment. A stem cell niche is a microenvironment in the extracellular matrix in which a stem cell resides. The extracellular matrix components affect growth, differentiation and mobility of stem cells through a multitude of peptide regulators, adhesions molecules, growth factors and transcriptional factors [14]. Among these regulators, small regulatory peptides, often described under an umbrella term “matrikines,” attract particular interest [15].

Matrikines (or matrycryptins) are small peptides that are released from a tissue after an injury, during an enzymatic cleavage of tissue proteins. Matrikines are attractive candidates for future anti-cancer and tissue regeneration-enhancing drugs; however, the application is hindered by the complexity of their interaction [16]. One of the potentially promising approaches, in our opinion, is identifying molecules that have “the master-key” qualities. One of the most promising candidates is the copper-binding peptide GHK (glycine-L-histidine-lysine) [17]. This peptide discovered in 1973 by Pickart, has the highest concentration in young adults (age 25 and younger), when tissues have the most regenerative potential; however, it declines with age [18].

Early studies by Maquart et al. established that GHK stimulates wound healing and tissue regeneration, increases collagen, elastin and glycosaminoglycans. Since then, multiple animal and in vitro studies confirmed GHK-Cu’s ability to improve wound healing and tissue regeneration [19, 20]. The positive effects of GHK-Cu were confirmed for many organs and systems such as skin, lungs, liver, intestinal lining, nervous system and bones. The effects of GHK cover a wide range of physiological processes, from regeneration and wound healing to anxiolytic, anti-aggression and analgesic effects [21-25]. GHK increases the level of antioxidant enzymes and has an anti-inflammatory effect [26]. The effects of GHK-Cu are summarized in Table 1.

The real breakthrough happened in 2010, when Hong at al. used the Broad Institute gene response database The Connectivity Map to identify the top molecules capable of reversing gene expression characteristic for metastatic colon cancer. Two molecules, GHK and securinine topped the list and were selected out of 1,309 bioactive molecules as the best agents capable to reverse expression of 54 gene sets overexpressed in malignant invasive metastatic colon cancer, which included node molecules YWHAB, MAP3K5, LMNA, APP, GNAQ, F3, NFATC2, and TGM2, involved in regulation of multiple biochemical pathways [27]. Surprisingly, both molecules happened to also

be stimulators of tissue regeneration. This study brought attention to GHK’s ability to modulate expression of multiple genes relevant to cancer suppression and tissue regeneration.

Table 1. Biological effects of GHK-Cu

Cell, tissue, organ	Effects	Study comments
Skin	Reduces wrinkles, improves complexion, reduces mottled pigmentation, improves wound healing, increases antioxidant enzymes, reduces inflammation, regulates remodeling of dermal matrix, stimulates collagen and elastin and other components of dermal matrix, regulates activity of metalloproteases.	Cosmetic studies, wound healing studies (rats, mice, rabbits, dogs), cultured fibroblasts and keratinocytes, engineered skin
Basal skin cells	Stimulates stemness, increases p53 and integrins.	Engineered skin
Fibroblasts	Stimulates collagen, elastin, decorin and glycosaminoglycan synthesis, restores functionality of irradiated fibroblasts.	Wound healing studies (rat wound chambers), cell cultures.
Hair	Improves hair growth, improves survival of hair grafts.	Animal studies (mice), hair grafting, hair follicles.
Bone	Stimulates regeneration	Animal studies (rats), cell culture.
Liver	Protects from damage, improves functionality of liver cells, reverts synthesis of proteins to that compared to younger liver cells.	Animal studies (rats), cell culture.
Nervous system	Reduces pain and anxiety, stimulates synthesis of neurotrophic factors.	Animal studies (anti-anxiety and anti-pain effects in rats), tissue and cell culture.
Intestine	Repairs damage of intestine and stomach lining, reduces inflammation.	Animal studies (rats).
Lungs	Reverts gene signature of COPD lungs to healthy expression profile. Switches regulation from destruction to repair. Protects from lipopolysaccharide induced damage	Gene profiling studies, animal studies (mice), human tissue and cell culture.

In 2012, Campbell et al. demonstrated that GHK can reverse gene expression characteristic for COPD (Chronic Obstructive Lung Disease). This study is of particular interest because the researchers also paired gene studies with laboratory studies, confirming that changes in gene expression are associated with improved collagen reorganization and fibroblasts function [28] Further studies confirmed the ability of GHK-Cu to reverse gene expression in COPD back to health

[29]. These studies provided an intriguing link between GHK's role in cancer prevention, tissue regeneration and stem cells.

The current paper reviews GHK's effect on gene expression relevant to stem cell function and regeneration.

2. Materials and Methods

The authors used the Broad Institute's Connectivity Map (CMap) - a publicly available computer-based gene profiling tool. The Connectivity Map is a database that contains more than 7,000 gene expression profiles of 5 human cell lines treated with 1,309 distinct small molecules [30].

The GHK profiles, contained in this repository, are based on PC3 human prostate cancer cells and MCF7 human breast cancer cells. In order to analyze the gene data obtained from the CMap, we used GenePattern - a publicly available computational biology open-source software package developed for the analysis of genomic data. The CEL files were processed with MAS5 and background correction. Files were then uploaded to the ComparativeMarkerSelectionViewer module in order to view fold changes for each probe set. The Gene Ontology descriptions of the molecular function, biological process, and cellular component of gene products was used to obtain GHK induced gene expression results relevant to stem cells.

Due to multiple probe sets mapping to the same gene, we converted the fold changes in mRNA production produced by GenePattern to percentages, then averaged all probe sets representing the same gene. It was determined that the 22,277 probe sets in the Broad data represent 13,424 genes. This ratio (1.66) was used to calculate the overall number of genes that affect GHK at various cutoff points (rather than probe sets). This method has been described in our previous publications, where we presented our data on genes relevant to nervous system function, antioxidant system, ubiquitin-proteasome system, skin health and tissue regeneration affected by GHK [31-34]. This paper presents gene profiling data relevant to GHK's effects on stem cells function.

It is important to notice that The Broad Institute gene profiling was performed with GHK, not GHK-Cu. However, considering GHK's high affinity for copper (see below) and essentiality of copper for cell growth, it is virtually impossible to have copper-free GHK in a living system. If ample copper 2+ is available in a test system or an animal, GHK should easily obtain necessary copper.

3. Results

The number of human genes associated with stem cell function was 57 genes in the range of increases of 50% UP and 46 genes in the range of decreases of 50% DOWN. Table 2 summarizes GHK's effects on genes relevant to cell cycle regulation. Table 3 presents genes that are upregulated, and Table 4 presents genes that are down regulated.

Table 2 Genes relevant to cell cycle regulation upregulated by GHK

Gene Abbreviation and Name (GENE database)	Gene Function (Gene Database)	Percentage of change in expression (The Broad Institute cMAP)
USP 29 Ubiquitin specific peptidase 29	Increases accumulation of p53	+1053
TP73 tumor protein p73	A member of p53 family of transcriptional factors.	+ 938
CAS 1 Caspase 1	Participates in regulations of multiple cellular pathways, including those involved in cell motility, apoptosis and cell cycle control.	+ 432
RAD50 Double strand break repair protein	DNA repair, cell cycle checkpoint activation.	+ 175
PTEN Phosphatase and tensin homolog	Cancer suppressor gene	+165
TGB2 transforming tumor factor beta receptor 2	This receptor/ligand complex phosphorylates proteins, which then enter the nucleus and regulate the transcription of genes related to cell proliferation, cell cycle arrest, wound healing, immunosuppression, and tumorigenesis.	+129

Table 3 Genes relevant to stem cell function upregulated by GHK

Gene Abbreviation and Name (GENE database)	Gene Function (Gene Database)	Percentage of change in expression (The Broad Institute cMAP)
MECOM, MDS1 and EVI1 complex locus	Critical for long-term hematopoietic stem cell function.	+578
SMO, smoothed, fizzled class receptor	G protein-coupled receptor that interacts with the patched protein, a receptor for hedgehog proteins.	+415
ESRRB, estrogen-related receptor beta	A similar protein in mice plays role in placenta development	+415
MED12, Mediator complex subunit 12	Participates in transcription regulation.	+393
WNT3, Wnt family member 3	A member of the WNT gene family. Plays role in several developmental processes, including regulation of cell fate and patterning during embryogenesis.	+355

ING2, Inhibitor of growth family, member 2	Function in regulation of DNA repair and apoptosis	+337
ZBTB16, zinc finger and BTB domain containing 16	Involved in cell cycle progression.	+299
GPLD1, glycosylphosphatidylinositol specific phospholipase D1	Encodes an enzyme that releases attached proteins from the plasma membrane.	+290
jagged 1, JAG1	Plays role in hematopoiesis.	+222
TAL1 TAL bHLH transcription factor 1, erythroid differentiation factor	Plays role in regulation of blood development.	+215
WNT2B, Wnt family member 2B	WNT family members function in a variety of developmental processes including regulation of cell growth and differentiation.	+190
APC, WNT signaling pathway regulator	A tumor suppressor protein that acts as an antagonist of the Wnt signaling pathway. Plays role in cell migration and adhesion, transcriptional activation, and apoptosis.	+204
GATA2, GATA binding protein 2	Plays an essential role in regulating transcription of genes involved in the development and proliferation of hematopoietic and endocrine cell lineages.	+195
POU5F1P4, POU class 5 homeobox 1 pseudogene 4	Transcriptional activator, eye development.	+193
SOX11, SRY (sex determining region Y)-box 11	A member of the SRY-related HMG-box (SOX) family of transcription factors involved in the regulation of embryonic development and in the determination of cell fate.	+154
GPM6A, glycoprotein M6A	Involved in neuronal differentiation and migration of neuronal stem cells.	+140
RUNX1T1, runt-related transcription factor 1, (cyclin D-related)	A member of the myeloid translocation gene family which interact with DNA-bound transcription factors and recruit a range of corepressors to facilitate transcriptional repression.	+140
SMARCA4, SWI/SNF related, matrix associated, actin dependent regulator of	Thought to regulate transcription of certain genes by altering the chromatin structure around those	+136

chromatin, subfamily a, member 4	genes.	
TGFB2, transforming growth factor beta 2	Recruitment and activation of SMAD transcription factors.	+129
SRRT, serrate, RNA effector molecule	Plays a central role in RNA recognition and processing.	+125
NOTCH2, Notch 2	Notch family members play a role in a variety of developmental processes by controlling cell fate decisions.	+123
MED6, mediator complex subunit 6	Involved in transcription of polymerase II genes.	+106
RUNX1, runt-related transcriptional factor 1	Plays role in hemopoiesis.	+103

Table 4 Gene relevant to stem cells function suppressed by GHK

Gene Abbreviation and Name (GENE database)	Gene Function (Gene Database)	Percentage of change in expression (The Broad Institute cMAP)
ABCB1, ATP binding cassette subfamily B member 1	ABC proteins transport various molecules across extra- and intra-cellular membranes.	-1536
FGFR2, fibroblast growth factor receptor 2	A member of the fibroblast growth factor receptor family, involved in regulation of cell growth and differentiation.	-337
ACE, angiotensin I converting enzyme (peptidyl-dipeptidase A)	Controls blood pressure.	-314
PTPRC, protein tyrosine phosphatase, receptor type C	A member of a family of proteins that regulate a variety of cellular processes including cell growth, differentiation, mitosis, and oncogenic transformation.	-336
TACSTD2, tumor associated calcium signal transducer 2	This intronless gene encodes a carcinoma-associated antigen.	-306
MCPH1, microcephalin 1	The encoded protein may play a role in G2/M checkpoint arrest	-275
MED14, mediator complex subunit 14	Participates in transcriptional activation	-262
LIN28A, lin-28 homolog A	A LIN-28 family RNA-binding protein that acts as a posttranscriptional regulator of genes involved in developmental timing and self-	-259

	renewal in embryonic stem cells.	
NGF, nerve growth factor	Stimulation of nerve growth	-242
SHH, sonic hedgehog	Plays crucial role in embryonic development	-220
PIWIL2, piwi like RNA-mediated gene silencing 2	Function in development and maintenance of germline stem cells	-214
PDGFRA, platelet-derived growth factor receptor, alpha polypeptide	A cell surface tyrosine kinase receptor for members of the platelet-derived growth factor family. These growth factors are mitogens for cells of mesenchymal origin.	-195
CNOT2, CCR4-NOT transcription complex subunit 2	Regulates mRNA synthesis and degradation and is also thought to be involved in mRNA splicing, transport and localization	-166
SCT, secretin	A member of glucagon family of peptides.	-125
TP63, tumor protein 63	A member of the p53 family of transcription factors.	-117
SOX21, SRY (sex-determining region Y) box 21	A member of the SRY-related HMG-box (SOX) family of transcription factors involved in the regulation of embryonic development and in the determination of cell fate.	-110
PDX1, pancreatic and duodenal homeobox 1	Affects insulin, somatostatin, glucokinase, islet amyloid and glucose.	-101
A2M, alpha-2-macroglobulin	An antiprotease, able to inactivate a variety of proteinases.	-101

4. Discussion

4.1 GHK and Gene Expression Relevant to Stem Cell Function

An examination of the GHK-induced actions (Tables 2-4) on gene expression relevant to stem cell function finds many genes that control development and differentiation, cell growth, RNA and DNA synthesis and transcription. For some genes, the Broad Institute database gives more than one value and they are averaged in the tables.

In some cases, it appears that GHK induces a selection pattern among genes with similar functions. For example, GHK suppresses gene ACBC1 (ATP-binding Cassette) by 1,536% but only suppresses a similar gene ACCB4 by only 70%. Likewise, gene ING2, which functions in DNA repair and apoptosis, is increased by 336% while gene PIWIL2, which eliminates germ lines with DNA damage is suppressed by 214 %. Also, there is the problem that the function of many genes is still

poorly defined.

It is known that GHK is a normal constituent of human plasma and may be a breakdown product of collagen. In addition, it is generated from SPARC (secreted protein acidic and rich in cysteine). The expression of SPARC is minimal in most normal adult tissues but is increased after injury. Proteolysis of this protein by plasmin generates GHK [35].

We propose that during wound repair, the initially generated GHK is copper free and stimulates stem cell replication. Later, as GHK accumulates bound copper 2+, GHK-Cu stimulates stem cell differentiation.

4.2 GHK and Stem Cells

The first suggestion that GHK might induce stem cell production arose from wound healing studies. Burn surgeons have long observed that the influx of hair follicles into a burned area predicts a good healing response. It is now established that dermal hair follicles provide a major source of stem cells used for dermal healing. Stem cells for the skin are thought to arise from enlarged hair follicles. The first indication that GHK affects stem cells came from mouse studies where GHK-Cu produced a very strong amplification of hair follicle size (see the description below). A similar peptide, Ala-His-Lys-copper 2+, produced even stronger actions [36].

The mouse in Figure 1 was shaved, then treated in three spots with GHK-Cu. The result is a much more rapid hair growth (the three circular patches of hair) in the spots treated with copper peptides. In the microscopic images, the magnifications are identical. The top photo is the untreated mouse skin – the control. The bottom photo is mouse skin treated with GHK-Cu. Note the larger hair follicles (elongated columns) in the lower photo, the increased content of subcutaneous fat in the skin (white material in the center of the skin), and the increased thickness of the skin. Hair researchers have noted the accumulation of such fat around healthy follicles that are vigorously growing hair and its relative lack around dormant follicles. They have postulated that these cells serve a supportive function for the hair follicle. It must be emphasized that effects in humans on hair follicle health are not as dramatic.

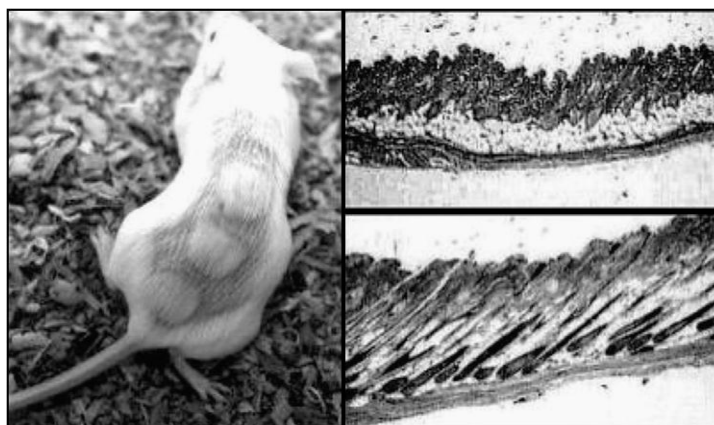


Figure1 GHK-Cu was tested for effects of hair growth in 30-day old Swiss-Webster mice. Control mice were treated with 0.85% saline in water. On the left: A mouse was shaved and treated with GHK-Cu on three areas. On the right top: Control - 0.85% saline-treated skin. On the right bottom: GHK-Cu treated skin.

4.3 GHK-Cu's Possible Mechanism of Action

Despite decades of research, there is still a lack of unanimous agreement among researchers as to what allows GHK to produce so many diverse and positive biological effects. In 1980, Pickart proposed that GHK acts by regulating copper intake into the cell [37]. Copper is an essential element in the human body, and it is involved in multiple biochemical processes which affect cell growth and differentiation, tissue regeneration, nervous system health and development, and gene expression [38]. Since then, biological effects of GHK have been commonly attributed to its role in copper metabolism.

There are approximately 700 albumin molecules in human plasma for each GHK molecule, so GHK does not play a significant role in copper 2+ transport. However, GHK has some unique characteristics which may be attributed to its importance as a copper-regulating compound on a cellular level. It has been established that GHK has a very high affinity for copper 2+ and can easily obtain the ionic metal from its transport site on human albumin. A comparison of copper binding properties, copper exchange rate and redox potential between two similar naturally occurring copper-binding peptides GHK and DAHK shows that even though both bind copper in stoichiometry at a ratio of 1:1, the exchange rate of copper between DAHK peptides is very slow, while the exchange rate between GHK peptides is much faster. Both complexes are inert under moderate redox conditions and do not increase oxidation [39]. Due to its small size and unique copper-binding characteristics, GHK may be able to facilitate rapid exchange of copper in the intracellular space.

Even though, considering the essentiality of copper in humans, the copper hypothesis of GHK mechanism of action seems very attractive, recent gene data may challenge this mechanism and demand more studies into GHK's mechanism of action.

4.4 GHK, Copper and Stem Cells

Stem cell proliferation requires extremely low copper concentrations that are created by the use of copper chelating agents. However, when stem cells are exposed to higher copper levels, they progress into differentiated cells. Peled and colleagues, members of a stem cell biotech firm (Gamida Cell, Jerusalem, Israel), has claimed, in a patent, that GHK increases proliferation of stem cells, while GHK-Cu increases their progression into differentiated cells [40]. In 2005, Peled and colleagues claimed that copper-free GHK maintained the clonogenic potential of stem cells while GHK-Cu increased cell copper by 2,162 % above the control value and caused stem cell differentiation [41].

Further supporting the above are the findings that low tissue copper, in itself, may increase stem cell proliferation and availability in animals. For instance, copper deprivation contributes to neogenesis of alpha and beta cells in the pancreatic ducts of diabetic rats [42].

Feeding diabetes-prone BioBreeding (BBdp) rats a hydrolyzed casein-based diet, a diet that binds nutritional copper and lowers tissue copper, promotes islet cell neogenesis and results in 2–3-fold fewer diabetes cases compared with feeding cereal-based diets [43].

Jose et al. found that the human plasma copper binding peptide GHK added to cultures of mesenchymal stem cells increased their concentrations of VEGF and basic fibroblast growth factor,

and also increased endothelial cell proliferation, migration and tubule formation. GHK also had no apparent cytotoxic effects on MSC in culture over a wide range of concentrations [44].

4.5 Anti-inflammatory Activity of GHK-Cu

As stated above, inflammation is one of the factors that can disrupt wound healing and trigger abnormal cell plasticity. There is an ample evidence that GHK-Cu possesses a strong anti-inflammatory effect.

GHK administered intraperitoneally at doses of 2.6, 26, and 260 µg/ml/day reduced inflammatory cell infiltration, levels of TNF-α and IL-6, as well as abolished bleomycin-induced elevation of TGF-beta in bleomycin-induced lung fibrosis in mice. The authors concluded that GHK acts by affecting TGF-β1/Smad 2/3 and IGF-1 pathway. [45].

GHK protected lungs from lipopolysaccharide-induced damage by suppressing infiltration of inflammatory cells into lungs and reducing inflammatory response. It also suppressed NF-κB p65 and p38 MAPK signaling pathways and protected the lungs by reducing reactive oxygen species (ROS) production, stimulating superoxide dismutase (SOD), while decreasing TNF-α and IL-6 production [46].

Also, GHK and GHK-Cu decreased TNF-alpha-dependent secretion of pro-inflammatory IL-6 in normal human dermal fibroblasts NHDF cell line [47].

4.6 DNA Repair

In our previous publications [29], we identified genes relevant to DNA repair that were positively affected by GHK (Table 5).

Table 5 Genes relevant to DNA repair affected by GHK

Gene title (GENE database)	Comments	Percent change in gene expression
PARP3, Poly (ADP-ribose) polymerase family, member 3	PARP3 enzymes play role in DNA repair, regulation of apoptosis, and maintenance of genomic stability	+253
POLM, Polymerase (DNA directed), mu	DNA repair enzyme	+225
RAD50, RAD50 double strand break repair protein	DNA repair enzyme, double strand break repair	+175
RARA, Retinoic acid receptor, alpha	Regulates transcription	+123

4.7 GHK vs GHK-Cu

There are many studies that obtain significant results with copper-free GHK, but the copper chelator bathocuproine abolishes the GHK effects. It is difficult to remove trace amounts of copper ions from the amino acids mixtures used in cell culture media.

Two studies from Seoul National University (Republic of Korea) found the effects of GHK-Cu and GHK copper-free on cell differentiation on a skin equivalent cell culture system produced identical

results. Both molecules (0.1–10 μM) stimulated the proliferation of keratinocytes in a dose dependent manner and resulted in changes in epidermal basal cells whose integrins and p63 expression markedly increased. Epidermal cells became more cuboidal, as is characteristic for stem cells. The authors concluded that GHK and GHK-Cu increased epidermal cell stemness and keratinocyte proliferation, also increasing p63 and integrin expression [48, 49].

In most of our animal studies, we used a mixture of 2 molecules of GHK to 1 molecule of copper ion to avoid the possible oxidative actions of free copper ion.

5. Conclusions

Recently, many compounds, discovered through computer-based gene-profiling, were proposed as novel therapeutics. In our opinion, when considering a new compound for clinical testing, it is important to have in vivo and in vitro studies confirming positive effects suggested by gene profiling as well as safety. GHK-Cu is a compound which has a long history of safe usage in skin care and which has been studied extensively in animal experiments, and in cell and tissue culture studies. Recent gene profiling studies allow deeper understanding of mechanisms of its positive actions.

Complex influences in stem cells' microenvironment determine whether the stem cell will function normally, improving tissue repair and regeneration, or transform into a malignant cell. Stem cell senescence also depends on gene expression and microenvironmental cues. Both laboratory and gene studies suggest that GHK, a very safe molecule and low-cost molecule, could improve stem cell therapies.

The Lethal Dose of GHK-Cu for 50% of mice (LD50) deaths was 8 mg for a 25-gram mouse or about 23 grams for a 70 kg human. A possible explanation of lethal effects of high doses of GHK-Cu is dropping of blood pressure. There are no reports for an LD50 for GHK without copper, so it must be very nontoxic. However, penicillamine, a copper chelator used in Wilson's Disease, has been reported to, at times, both induce psychosis or reverse psychosis.

For experimental human use, dissolved GHK could be added to the stem cell mixture. A total of 10 mg in an adult human should be adequate, which is about 2000 times lower than the LD50 calculated for humans. Recent Russian studies of rats observed a reduction of anxiety and pain induced aggression 12 minutes after intraperitoneal administration of GHK at concentrations that would correspond to 35 micrograms in a 70 kg human.

To improve skin regeneration GHK can be used in creams, gels and skin patches, since it can penetrate the stratum corneum [50]. It can be delivered directly into the affected site through the use of microneedles [51]. Also, GHK can be easily incorporated in liposomes and used as a dietary supplement [52].

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Author Contributions

Authors equally contributed to research and preparation of the manuscript.

Competing Interests

The authors have declared that no competing interests exist.

References

1. Blanpain C, Fuchs E. Stem Cell Plasticity. Plasticity of epithelial stem cells in tissue regeneration. *science*. 2014; 13: 1242281. doi: 10.1126/science.1242281
2. Gonzales KAU, Fuchs E. Skin and its regenerative powers: an alliance between stem cells and their niche. *Dev Cell*. 2017; 20: 387–401.
3. Milanovic M, Fan DNY, Belenki D, Däbritz JHM, Zhao Z, Yu Y, et al. Senescence-associated reprogramming promotes cancer stemness. *Nature*. 2018; 553: 96-100.
4. Dou Z, Berger SL. Senescence elicits stemness: a surprising mechanism for cancer relapse. *Cell Metab*. 2018; 27: 710-711.
5. Ritschka B, Storer M, Mas A, Heinzmann F, Ortells MC, Morton JP, et al. The senescence-associated secretory phenotype induces cellular plasticity and tissue regeneration. *Genes Dev*. 2017; 15: 172-183.
6. Ortiz-Montero P, Londoño-Vallejo A, Vernot JP. Senescence-associated IL-6 and IL-8 cytokines induce a self- and cross-reinforced senescence/inflammatory milieu strengthening tumorigenic capabilities in the MCF-7 breast cancer cell line. *Cell Commun Signal*. 2017; 15: 17. doi: 10.1186/s12964-017-0172-3.
7. Arnold KM, Opdenaker LM, Flynn D, Sims-Mourtada J. Wound healing and cancer stem cells: inflammation as a driver of treatment resistance in breast cancer. *Cancer Growth Metastasis*. 2015; 8: 1-13.
8. Yang Z, Balic A, Michon F, Juuri E, Thesleff I. Mesenchymal wnt/ β -Catenin signaling controls epithelial stem cell homeostasis in teeth by inhibiting the antiapoptotic effect of fgf10. *Stem Cells*. 2015; 33: 1670-1681.
9. Brum AM, Van de Peppel J, Nguyen L, Aliev A, Schreuders-Koedam M, Gajadien T, et al. Using the connectivity map to discover compounds influencing human osteoblast differentiation. *J Cell Physiol*. 2018; 233: 4895-4906.
10. Meng S, Chanda P, Thandavarayan RA, Cooke JP. Transflammation: how innate immune activation and free radicals drive nuclear reprogramming. *Antioxid Redox Signal*. 2018; 29: 205-218.
11. Liu X, Grogan TR, Hieronymus H, Hashimoto T, Mottahedeh J, Cheng D, et al. Identifies progenitor-like inflammation-associated luminal cells that can initiate human prostate cancer and predict poor outcome. *Cell Rep*. 2016; 17: 2596-2606.
12. Perochon J, Carroll LR, Cordero JB. Wnt signalling in intestinal stem cells: lessons from mice and flies. *Genes (Basel)*. 2018; 9: 138.
13. Olivos DJ, Mayo LD. Emerging non-canonical functions and regulation by p53: p53 and stemness. *Int J Mol Sci*. 2016; 17: 1982.
14. Gattazzo F, Urciuolo A, Bonaldo P. Extracellular matrix: a mynamic microenvironment for stem cell niche. *Biochim Biophys Acta*. 2014; 1840: 2506-2519.

15. Gaur M, Dobke M, Lunyak VV. Mesenchymal stem cells from adipose tissue in clinical applications for dermatological indications and skin aging. *Int J Mol Sci.* 2017; 18. doi:10.3390/ijms18010208.
16. Ricard-Blum S, Vallet SD. Matricryptins network with matricellular receptors at the surface of endothelial and tumor cells. *Front Pharmacol.* 2016; 7. doi: 10.3389/fphar.2016.00011.
17. Maquart FX, Siméon A, Pasco S, Monboisse C. Regulation of cell activity by the extracellular matrix: the concept of matrikines. *J Soc Biol.* 1999; 193: 423-428.
18. Pickart L. A Tripeptide from human serum which enhances the growth of neoplastic hepatocytes and the survival of normal hepatocytes. San Francisco: University of California; 1973.
19. Pickart L, Margolina A. Anti-aging activity of the GHK peptide—the skin and beyond. *J Aging Res Clin Pract.* 2012; 1: 13–16.
20. Badenhorst T, Svirskis D, Merrilees M, Bolke L, Wu, Z. Effects of GHK-Cu on MMP and TIMP expression, collagen and elastin production, and facial wrinkle parameters. *J Aging Sci.* 2016; 4. doi: 10.4172/2329-8847.1000166.
21. Bobyntsev II, Chernysheva OI, Dolgintsev ME, Smakhtin MY, Belykh AE. Anxiolytic effects of Gly-His-Lys peptide and its analogs. *Bull Exp Bio Med.* 2015; 158: 726-728.
22. Sever'yanova LA, Dolgintsev ME. Effects of tripeptide Gly-His-Lys in pain-induced aggressive-defensive behavior in rats. *Bull Exp Bio Med.* 2017; 164: 140-143.
23. Bobyntsev II, Chernysheva OI, Dolgintsev ME, Smakhtin M, Belykh AE. Effect of Gly-His-Lys peptide and its analogs on pain sensitivity in mice. *Eksp Klin Farmakol.* 2015; 78: 13-15.
24. Chernysheva OI, Bobyntsev II, Dolgintsev ME. The tripeptide GLY-HIS-LYS influence on behavior of rats in the test “open field”. *Biological Sci.* 2014; 12: 357-360.
25. Pickart L, Vasquez-Soltero JM, Margolina A. Resetting skin genome back to health naturally with GHK. *Textbook of Aging Skin, 2014: 1549-1566.*
26. Pickart L, Vasquez-Soltero JM, Margolina A. GHK-Cu may prevent oxidative stress in skin by regulating copper and modifying expression of numerous antioxidant genes. *Cosmetics,* 2015; 2: 236-247.
27. Hong Y, Downey T, Eu KW, Koh PK, Cheah PY. A ‘metastasis-prone’ signature for early-stage mismatch-repair proficient sporadic colorectal cancer patients and its implications for possible therapeutics. *Clinical & Exp Metastasis.* 2010; 27: 83-90.
28. Campbell JD, McDonough JE, Zeskind JE, Hackett TL, Pechkovsky DV, Brandsma CA, et al. A gene expression signature of emphysema-related lung destruction and its reversal by the tripeptide GHK. *Genome Med.* 2012; 4: 67. doi: 10.1186/gm367.
29. McMurry R, Perdomo C, Liu G, Zhang S, Stevenson C, Campbell J, et al. GHK-Cu elicits in vitro, dose-dependent transcriptional alterations in pathways relevant to extracellular matrix composition. *Am J Resp Critical Care Medicine,* 2017; 195: A2446.
30. Lamb J. The connectivity map: a new tool for biomedical research. *Nat Rev Cancer.* 2007; 7: 54-60.
31. Pickart L, Vasquez-Soltero JM, Margolina A. The effect of the human peptide GHK on gene expression relevant to nervous system function and cognitive decline. *Brain Sci.* 2017; 7. doi: 10.3390/brainsci7020020.
32. Pickart L, Vasquez-Soltero JM, Margolina A. GHK and DNA: resetting the human genome to health. *BioMed Res Int.* 2014; 151479. doi: 10.1155/2014/151479.

33. Pickart L, Margolina A. Regenerative and protective actions of the GHK-Cu peptide in the light of the new gene data. *Int J Mol Sci.* 2018; 19. doi: 10.3390/ijms19071987.
34. Pickart L, Vasquez-Soltero JM, Margolina A. GHK peptide as a natural modulator of multiple cellular pathways in skin regeneration. *BioMed Res Int.* 2015; 648108. doi: 10.1155/2015/648108
35. Sage EH, Vernon RB. Regulation of angiogenesis by extracellular matrix: the growth and the glue. *J Hypertens Suppl.* 1994; 12: 145-152.
36. Pickart L. The human tri-peptide GHK and tissue remodeling. *J Biomaterials Sci.* 2008; 19: 969-988.
37. Pickart L, Freedman JH, Loker WJ, Peisach J, Perkins CM, Stenkamp RE, et al. Growth-modulating plasma tripeptide may function by facilitating copper uptake into cells. *Nature.* 1980; 288: 715-717.
38. Uauy R, Olivares M, Gonzalez M. Essentiality of copper in humans. *Am J Clin Nutr.* 1998; 67: 952-959.
39. Hureau C, Eury H, Guillot R, Bijani C, Sayen S, Solari PL, et al. X-ray and solution structures of Cu(II) GHK and Cu(II) DAHK complexes: influence on their redox properties. *Chemistry.* 2011; 17: 10151-10160.
40. Peled T, Landau E, Prus E, Treves AJ, Fibach E. Cellular copper content modulates differentiation and self-renewal in cultures of cord blood-derived CD34+ Cells. *Br J Haematol.* 2002; 116: 655-661
41. Peled T, Fibach E, Treves A. U.S. Patent No. 7,855,075. Washington: Patent and Trademark Office. 2010.
42. Al-Abdullah IH, Ayala G, Panigrahi D, Kumar A M, Kumar MSA. Neogenesis of pancreatic endocrine cells in copper-deprived rat models. *Pancreas.* 2000; 21: 63-68.
43. Wang GS, Gruber H, Smyth P, Pulido O, Rosenberg L, Duguid W, et al. Hydrolysed casein diet protects BB rats from developing diabetes by promoting islet neogenesis. *J Autoimmun.* 2000; 15: 407-416.
44. Jose S, Hughbanks ML, Binder BY, Ingavle GC, Leach JK. Enhanced trophic factor secretion by mesenchymal stem/stromal cells with Glycine-Histidine-Lysine (GHK)-modified alginate hydrogels. *Acta Biomater.* 2014; 10: 1955-1964.
45. Zhou XM, Wang GL, Wang XB, Liu L, Zhang Q, Yin Y, et al. GHK peptide inhibits bleomycin-Induced pulmonary fibrosis in mice by suppressing TGF β 1/Smad-mediated epithelial-to-mesenchymal transition. *Front Pharmacol.* 2017; 904. doi: 10.3389/fphar.2017.00904.
46. Park JR, Lee H, Kim SI, Yang SR. The tri-peptide GHK-Cu complex ameliorates lipopolysaccharide-induced acute lung injury in mice. *Oncotarget.* 2016; 7: 58405-58417.
47. Gruchlik A, Jurzak M, Chodurek E, Dzierzewicz Z. Effect of Gly-Gly-His, Gly-His-Lys and their copper complexes on TNF-alpha-dependent IL-6 Secretion in normal human dermal fibroblasts. *Acta Pol Pharm.* 2012; 69: 1303-1306.
48. Kang YA, Choi HR, Na JI, Huh CH, Kim MJ, Youn SW, et al. Copper-GHK increases integrin expression and p63 positivity by keratinocytes. *Arch Dermatol Res.* 2009; 301: 301-306.
49. Choi HR, Kang YA, Ryoo SJ, Shin JW, Na JI, Huh CH, et al. Stem cell recovering effect of copper-free GHK in skin. *J Pept Sci.* 2012; 18: 685-690.

50. Hostynek JJ, Dreher F, Maibach HI. Human skin retention and penetration of a copper tripeptide in vitro as function of skin layer towards anti-inflammatory therapy. *Inflamm Res* 2010, 59: 983–988.
51. Li H, Low YS, Chong HP, Zin MT, Lee CY, Li B, et al. Microneedle-Mediated delivery of copper peptide through skin. *Pharm Res.* 2015; 32: 2678-2689
52. Erdem S, Türkoglu M. Glycyl-L-Histidyl-L-Liysine-Cu(2+) loaded liposome formulations. *Marmara Pharm J.* 2010; 14: 91–97.



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